

Table 1. Molecular characterisation of chromosomal translocations in acute myeloid leukaemias

Disease subtype	% of all AML	Chromosomal translocation	Fusion product	Reference
1. Differentiated myeloid (FAB AML M2)	5–20	t(8;21)(q22;q22)	CBF $\alpha$ (AML1)-ETO	2
2. Myelomonocytic with eosinophilia (M4Eo)	8	inv (16)(p13;p32)	CBF $\beta$ -MYH 11	3
3. Promyelocytic (M3/M3v)	10	t(15;17)(q24;q21)	PML-RAR $\alpha$	4, 5
	Rare	t(11;17)(q23.1;q21)	PLZF-RAR $\alpha$	6
4. AML with basophilia	1–2	t(6;9)(p23;q34)	DEK-CAN	7

Fusion products deriving from translocation in AML generally represent the fusion of two transcription factors. Exceptions are MYH11 (smooth muscle myosin heavy chain) and RAR $\alpha$  (retinoic acid receptor- $\alpha$ ). PML, promyelocytic leukaemia gene; PLZF, promyelocytic zinc finger gene. Note that the frequency of the t(8;21) varies with geographical location.

PML/RAR $\alpha$  fusion, a condition analogous to chronic myeloid leukaemias which lack cytogenetically the t(9;22)(q34;q11), but have molecular evidence for BCR-ABL fusion [8]. Since clinical efficacy of ATRA depends on the expression of the PML/RAR $\alpha$  gene, evidence for this fusion product should be sought in all cases where such therapy is being considered.

The efficacy of ATRA in APL is a most exciting development. Much remains to be learnt, both clinically and scientifically. Might other forms of retinoic acid produce more durable CRs? Most importantly, do patients treated with ATRA fare as well as those treated with chemotherapy alone? (APL is one of the "best" subgroups of AML with regard to outcome.) Finally, given the experience in APL, do other translocations in other malignancies involve other RAR genes of which there are several, and if so, can the same approach to therapy be adopted?

1. Cleary ML. Transcription factors in human leukaemias. *Cancer Surv* 1992, 15, 89–104.

2. Miyoshi H, Shimizu K, Kozu T, Maseki N, Kaneko Y, Ohki M. t(8;21) breakpoints on chromosome 21 in acute myeloid leukaemia are clustered within a limited region of a single gene, AML1. *Proc Natl Acad Sci USA* 1991, 88, 10431–10434.
3. Liu P, Tarle SA, Hajra A, et al. Fusion between transcription factor CBF $\beta$ /PEBP2 $\beta$  and a myosin heavy chain in acute myeloid leukemia. *Science* 1993, 261, 1041–1044.
4. Warrell RP, De The H, Wang Z-Y, Degos L. Acute promyelocytic leukemia. *New Engl J Med* 1993, 329, 177–189.
5. Grinani F, Fagioli M, Alcalay M, et al. Acute promyelocytic leukemia: from genetics to treatment. *Blood* 1994, 83, 10–25.
6. Chen S-J, Zelent A, Tong J-H, et al. Rearrangements of the retinoic acid receptor alpha and promyelocytic zinc finger genes resulting from t(11;17)(q23;q21) in a patient with acute promyelocytic leukemia. *J Clin Invest* 1993, 91, 2260–2267.
7. Soekarman D, von Lindern M, van der Plas DC, et al. DEK-CAN rearrangement in translocation t(6;9)(p23;p34). *Leukemia* 1992, 6, 489–494.
8. Hiorns LR, Min T, Swansbury GJ, Zelent A, Dyer MJS, Catovsky D. Interstitial insertion of retinoic acid receptor alpha gene in acute promyelocytic leukemia with normal chromosomes 15 and 17. *Blood* 1994, in press.



Pergamon

European Journal of Cancer Vol. 30A, No. 5, pp. 574–576, 1994  
Elsevier Science Ltd  
Printed in Great Britain

0959-8049(93)E0127-P

## Serum Neurone-specific Enolase and Other Neuroendocrine Markers in Lung Cancer

J.A. Ledermann

SMALL CELL lung cancer (SCLC) can be distinguished from other types of lung cancer by its morphology, biological behaviour and chemosensitivity. Neuroendocrine differentiation is consistent, although not a specific feature of SCLC, and different patterns of neuroendocrine expression can be found in 'classic' and

'variant' subtypes of SCLC lines [1]. This raises the possibility that neuroendocrine markers might identify biological properties of SCLC which could provide useful clinical information.

Neurone-specific enolase (NSE), a neuronal form of the glycolytic enzyme enolase consistently found in SCLC lines, is a secreted biochemical marker of neuroendocrine tumours and SCLC [2]. The neuronal form of the enzyme contains either alpha-gamma or gamma-gamma subunits, and raised levels of NSE are found in the serum of 66–81% of patients with SCLC [3–5]. The detection of raised levels of NSE in serum correlates with its presence in tissue [6]. Elevated levels of other neuroendo-

Correspondence to J.A. Ledermann at the Department of Oncology, University College London Medical School, Middlesex Hospital, London W1N 8AA, U.K.

Received 22 Feb. 1994; accepted 2 Mar. 1994.

crine markers, such as creatine phosphokinase BB [7] or chromogranin A [8], may also be found in serum. Recently, another neuroendocrine marker, the cluster 1 antigen NCAM (neural cell adhesion molecule), has been added to the list. NCAM is present in high concentrations on the surface of SCLC [9]. There are several isoforms of NCAM, including a secreted product, and all of these may be found in SCLC tumours. NCAM levels are frequently elevated in the serum of patients with SCLC [10, 11].

Ideally, serum tumour markers should be diagnostic, correlate with disease extent, predict prognosis and influence therapeutic decisions. Only the embryonic tumour markers produced by germ cell malignancies fulfil all these criteria. Early detection of lung cancer is important as it may identify potentially curable disease, but so far there are no satisfactory screening methods. Attempts to screen for lung cancer by combining the results of measurements of four tumour markers, carcinoembryonic antigen, CA-50, tumour-specific trypsin inhibitor and NSE have not yielded encouraging results [12]. A diagnosis of lung cancer can usually be obtained by examination of sputum cytology, bronchoscopy or radiologically guided needle biopsy. Measurement of serum tumour markers may be useful when lung cancer is suspected but the diagnosis is uncertain. Bergman and colleagues [13] measured NSE, carcinoembryonic antigen and CA-50 in 168 patients with lung cancer and 102 patients with non-malignant chest conditions. Combining the results in a multivariate analysis, the diagnostic sensitivity of the tumour marker assays for lung cancer was 57% with a positive predictive value of 95%. In 22% of cases, a firm diagnosis of lung cancer was made more than 1 month after tumour markers were assayed. It is in this group that serodiagnosis could be most useful, but only half of these patients had previously had raised levels of tumour markers.

Therapeutic decisions in SCLC are usually made on the basis of complex and often costly staging procedures. These identify patients with widespread disease who have a worse prognosis. Serum NSE levels are generally higher in patients with 'extensive' compared with 'limited' stage disease [3, 4, 14, 15]. This is true for other neuroendocrine tumour markers, such as NCAM which correlates with NSE levels [10, 11]. However, measurement of serum NSE levels has not been shown to be able to predict reliably the presence of 'extensive' disease [16]. Furthermore, simple biochemical measurements and patient performance score provide an accurate prediction of prognosis [17], and these tests can be used to select patients who might benefit from more intensive treatment regimens. In a recent series, where serum NSE, chromogranin A and lactate dehydrogenase were measured in patients with SCLC NSE, performance status and serum albumin were shown by multivariate analysis to be the best independent predictors of survival. Twenty-five per cent of patients with NSE levels below twice the upper limit of normal, a Karnofsky score greater than 80 and a serum albumin greater than 35 g/l were alive at 2 years [5]. Jørgensen and colleagues [18] have shown in a multivariate analysis that disease-free survival was significantly prolonged in patients with 'limited' disease, those attaining a complete remission and those with pretreatment serum levels of NSE  $\leq 12.5$  ng/ml. Using a Cox regression analysis, they demonstrated that the probability of a complete response was related to disease extent but not NSE levels. However, NSE levels and the type of response were independent predictors of the duration of response. When examined as a continuous variable, NSE levels appear to relate to survival, with a reduction of approximately 10% in median

survival of patients with 'limited' disease for each 5 ng/ml incremental rise in NSE [19]. Drawing conclusions from all these studies, it seems that if complex staging is not performed to assess disease extent, pretreatment NSE levels might be useful in predicting disease-free survival.

As patients respond to treatment, levels of NSE fall; they are usually normal in patients in complete remission and often rise at relapse [3, 4]. Thus, NSE mirrors tumour bulk quite well, but in longitudinal studies, follow-up measurement of NSE has not been shown to be useful in predicting relapse [20, 21].

The neuroendocrine phenotype can also be demonstrated in some cases of non-small cell lung cancer (NSCLC), particularly adenocarcinoma, where features of neuroendocrine differentiation are found in 20–30% of cases [22–24]. Approximately 20% of patients with NSCLC have raised serum NSE levels [4]. There is some evidence that the presence of these markers may influence chemosensitivity or the metastatic potential of tumours. Two retrospective studies have shown an increased response rate to chemotherapy in patients with tumours expressing NSE [25, 26]. Serum NSE correlates with levels of lactate dehydrogenase, and both have been shown to be predictors of chemosensitivity [27]. However, studies have not shown that neuroendocrine differentiation, as an independent variable, is associated with a better or worse survival [24, 26, 28].

The literature on serum NSE in lung cancer is now quite extensive. The presence of neuroendocrine differentiation is used most often to support a diagnosis of SCLC. The reasons for the consistent expression of the neuroendocrine phenotype in this tumour and its association in some NSCLC remains enigmatic, but of great interest to researchers studying the biology of lung cancer. Measurement of NSE levels in NSCLC cannot be regarded as useful in routine clinical practice. However, in SCLC, knowledge of serum NSE values may help to predict recurrence-free survival.

1. Carney DN, Gazdar AF, Belper G, *et al.* Establishment and identification of small cell lung cancer cell lines having classic and variant features. *Cancer Res* 1985, **45**, 2913–2923.
2. Tapia FJ, Polak JM, Barbosa AJA, *et al.* Neurone specific enolase is produced by neuroendocrine tumours. *Lancet* 1981, **i**, 808–811.
3. Carney DN, Marangos PJ, Ihde DC, *et al.* Serum neuron-specific enolase: a marker for disease extent and response to therapy of small-cell lung cancer. *Lancet* 1982, **i**, 583–585.
4. Burghuber OC, Worofka B, Scherthaner G, *et al.* Serum neuron-specific enolase is a useful tumour marker for small cell lung cancer. *Cancer* 1990, **65**, 1386–1390.
5. Johnson PW, Joel SP, Love S, *et al.* Tumour markers for predication of survival and monitoring of remission in small cell lung cancer. *Br J Cancer* 1993, **67**, 760–766.
6. Jørgensen LGM, Hirsch FR, Skov BG, Østerlind K, Cooper EH, Larsson LI. Occurrence of neuron specific enolase in tumour tissue and serum in small cell lung cancer. *Br J Cancer* 1991, **63**, 151–153.
7. Carney DN, Zweig MH, Ihde DC, Cohen MH, Makuch RW, Gazdar AF. Elevated serum creatine kinase BB levels in patients with small cell lung cancer. *Cancer Res* 1984, **44**, 5399–5403.
8. Sobol RE, O'Connor T, Addison J, Suchocki K, Royston I, Deftos LJ. Elevated serum chromogranin A concentrations in small-cell lung carcinoma. *Ann Intern Med* 1986, **105**, 698–700.
9. Souhami RL, Beverley PCL, Bobrow LG. The antigens of small cell lung cancer. First international workshop. *Lancet* 1988, **ii**, 325–326.
10. Jaques G, Auerbach B, Pritsch M, Wolf M, Madry N, Havemann K. Evaluation of serum neural cell adhesion molecule as a new tumor marker in small cell lung cancer. *Cancer* 1993, **72**, 418–425.
11. Ledermann JA, Pasini F, Olabiran YO, Pelosi G. Detection of the neural cell adhesion molecule (NCAM) in serum of patients with small-cell lung cancer (SCLC) with "limited" or "extensive" disease, and bone marrow infiltration. *Int J Cancer* 1994, (suppl. 8), in press.

12. Järvisalo J, Hakama M, Knekt P, *et al.* Serum tumor markers CEA, CA 50, TATI, and NSE in lung cancer screening. *Cancer* 1993, **71**, 1982–1988.
13. Bergman B, Brizicka F-T, Engström C-P, Larsson S. Clinical usefulness of serum assays of neuron-specific enolase, carcinoembryonic antigen and CA-50 antigen in the diagnosis of lung cancer. *Eur J Cancer* 1993, **29A**, 198–202.
14. Johnson DH, Marangos P, Forbes JT, *et al.* Potential utility of serum neuron-specific enolase levels in small cell carcinoma of the lung. *Cancer Res* 1984, **44**, 5409–5414.
15. Cooper EH, Splinter TAW, Brown DA, Muers MF, Peake MD, Pearson SL. Evaluation of radioimmunoassay for neuron-specific enolase in small cell lung cancer. *Br J Cancer* 1985, **52**, 333–338.
16. Quoix E, Charloux A, Popin E, Pauli G. Inability to predict disease extent in small cell lung cancer based on initial level of serum neuron-specific enolase. *Eur J Cancer* 1993, **16**, 2248–2250.
17. Souhami RL, Bradbury I, Geddes DM, Spiro SG, Harper PG, Tobias JS. Prognostic significance of laboratory parameters measured at diagnosis in small cell carcinoma of the lung. *Cancer Res* 1985, **45**, 2878–2882.
18. Jørgensen LGM, Østerlind K, Hansen HH, Cooper EH. Serum neuron specific enolase (NSE) is a determinant of response duration in small cell lung cancer (SCLC). *Br J Cancer* 1992, **66**, 594–598.
19. Harding M, McAllister J, Hulks G, *et al.* Neurone specific enolase (NSE) in small cell lung cancer: a tumour marker of prognostic significance? *Br J Cancer* 1990, **61**, 605–607.
20. Nou E, Steinholtz L, Bergh J, Nilsson K, Pählman S. Neuron-specific enolase as a follow-up marker in small cell bronchial carcinoma. *Cancer* 1990, **65**, 1380–1385.
21. Van Zandwijk N, Jassem E, Bonfrer JMG, Van Tinteren H. Value of neuron specific enolase in early detection of relapse in small cell lung carcinoma. *Eur J Cancer* 1990, **26**, 373–376.
22. Bergh J, Esscher T, Steinholtz L, *et al.* Immunocytochemical demonstration of neuron-specific enolase (NSE) in human lung cancers. *Am J Clin Path* 1985, **84**, 1–7.
23. Dhillon AP, Rode J, Dhillon DP, *et al.* Neural markers in carcinoma of the lung. *Br J Cancer* 1985, **51**, 645–652.
24. Sundaresan V, Reeve JG, Stenning S, Stewart S, Bleehen NM. Neuroendocrine differentiation and clinical behaviour in non-small cell lung tumours. *Br J Cancer* 1991, **64**, 333–338.
25. Graziano SL, Mazid R, Newman N, *et al.* The use of neuroendocrine immunoperoxidase markers to predict chemotherapy response in patients with non-small-cell lung cancer. *J Clin Oncol* 1989, **7**, 1398–1406.
26. Skov BG, Sørensen JB, Hirsch FR, Larsson LI, Hansen HH. Prognostic impact of histologic demonstration of chromogranin A and neuron specific enolase in pulmonary adenocarcinoma. *Ann Oncol* 1991, **2**, 355–360.
27. van Zandwijk N, Jassem E, Bonfrer JM, Mooi WJ, Van Tinteren H. Serum neuron-specific enolase and lactate dehydrogenase as predictors of response to chemotherapy and survival in non-small cell lung cancer. *Semin Oncol* 1992, **19** (suppl. 2), 37–43.
28. Berendsen HH, de Leij L, Poppema S, *et al.* Clinical characterization of non-small-cell lung cancer tumours showing neuroendocrine differentiation features. *J Clin Oncol* 1989, **7**, 1614–1620.



Pergamon

European Journal of Cancer Vol. 30A, No. 5, pp. 576–577, 1994  
 Copyright © 1994 Elsevier Science Ltd  
 Printed in Great Britain. All rights reserved  
 0959-8049/94 \$7.00+0.00

0959-8049(93) E0080-A

## Radiosensitivity Testing of Normal Tissues: a Way to Optimise Radiotherapy?

P. Lambin and P. Lawton

IN THIS issue, Floyd and Cassoni (pp. 615–620) assessed the reliability of a lymphocyte micronucleus assay to determine the radiosensitivity of individual patients. Levels of radiation-induced micronuclei were measured following exposures of up to 4 Gy X-rays. The variation between individuals was greater than between repeat experiments on the same individual and, in accordance with the literature, they found that cord blood lymphocytes were generally more radiosensitive than normal lymphocytes. The authors conclude that the lymphocyte micronucleus assay could have some predictive capacity for the determination of individual radiosensitivity. Unfortunately, it is difficult to be certain as to whether or not the conclusions of Floyd and Cassoni are generally valid. Lymphocytes are a heterogeneous cell population, especially with regard to the proportion of lymphocyte subtypes. Furthermore, they die

after irradiation both by reproductive death and by apoptosis, in contrast to the majority of cell types which die mainly by a reproductive death. Despite these limitations, this is an important study. We urgently need a reliable assay for predicting the radiosensitivity of normal tissues before radiotherapy is commenced. In order to achieve this, it is essential to understand the relationship between the *in vitro* radiosensitivity of different cell types and the clinical response to radiotherapy.

### A subpopulation of radiosensitive patients

There is evidence to suggest that both tumours and normal tissues show interindividual differences in intrinsic radiosensitivity. In 1975, Taylor and colleagues noted a correlation between the cellular radiosensitivity of skin fibroblasts and severe reactions to radiotherapy in an individual with the genetic disorder ataxia teleangiectasia (AT) [1]. There have now been a number of published retrospective studies of individuals showing severe reactions to radiotherapy where cultured fibroblasts have shown excessive *in vitro* radiosensitivity [2–4].

AT is a rare genetic disorder but it has been estimated that about 1% of the population are carriers for the AT gene. These

Correspondence to P. Lambin at University Hospital St Rafaël, Department of Radiotherapy, Leuven, Belgium.

P. Lawton is at the Gray Laboratory, Mount Vernon Hospital, Northwood, Middlesex, U.K.

Received 23 Nov. 1993; accepted 9 Dec. 1993.